

# The Second International Conference on Genetic Resources and Biotechnology

## Harnessing Technology for Conservation and Sustainable Use of Genetic Resources for Food and Agriculture

Bogor, Indonesia • 24–25 May 2021

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January 2022

THE SECOND INTERNATIONAL CONFERENCE ON GENETIC RESOURCES  
AND BIOTECHNOLOGY: Harnessing Technology for Conservation and Sustainable  
Use of Genetic Resources for Food and Agriculture

# Committees: The Second International Conference on Genetic Resources and Biotechnology

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## **Preface: The Second International Conference on Genetic Resources and Biotechnology**

The Second International Conference on Genetic Resources and Biotechnology, which is the continuation of the first event held in 2018, focuses on topics related to advances in biotechnology to create more opportunities for effective conservation and sustainable utilization of genetic resources for food and agriculture. This year conference's theme is Harnessing Technology for Conservation and Sustainable Use of Genetic Resources for Food and Agriculture. The conference was organized by Indonesian Agency for Agricultural Research and Development (IAARD), Ministry of Agriculture, Indonesia, in collaboration with Indonesian Biotechnology Consortium and held on 24<sup>th</sup>-25<sup>th</sup> of May 2021 virtually due to the pandemic of COVID-19.

The conference aims to share and exchange current scientific information and technological developments on biotechnology and their applications for conservation and sustainable use of genetic, to encourage and promote quality, efficiency, and modernization of management and utilization of genetic resources, and to facilitate national and international collaboration among participants. There are five scopes discussed in this conference. They are effective management of conservation and sustainable use of genetic resources for food and agriculture, application of genomics and molecular markers for genetic resource conservation and crop adaptation to climate change, application of innovative crop improvement techniques for conservation and sustainable use of plant genetic resources for food and agriculture, plant cell and tissue culture for conservation and effective utilization of genetic resources, and the use of microbial genetic resources as biological control agents of agricultural pests and diseases, and for soil bioremediation.

Five speakers from the United States of America, Japan, India and Indonesia were invited to discuss about their expertise and knowledge on relevant subjects in the plenary sessions. This conference was attended by more than 100 participants including 75 presenters and 44 listeners worldwide. They came from diverse governmental, private, or academic institutions and also scientific communities. The presented materials have undergone peer review processes and only qualified papers were selected. Furthermore, all papers were subjected to double blind peer-review and expected to meet the scientific criteria of significance and academic excellence to be published in a conference proceedings indexed in a well-known, reputable service.

We would like to express our sincere gratitude to our speakers, presenters and all participants for their contributions in this conference. We would also like to express our appreciation for the generosity of our sponsors that support this conference: PT CropLife, PT ITS Science Indonesia, PT Fajar Mas Murni and PT Prima Instrument Analitika. Lastly, special thanks to all committee members for their exceptional work and contributions in the conference and publication.

Chair of Organizing Committee

Dr. Toto Hadiarto

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**Abstract.** Agroforensic will identify and solve the problem when one case such as genetic diversity of crops should be declared, and in other situation need some confirmation like as detection of genetic modified organism (GMO) on food product or agrobusiness. Identification of DNA fingerprinting in forensics can be presented as DNA pattern profile containing repetitive and varied sequences among the investigated individuals. Several PCR-based technologies have been used in such genetic analysis. This study was conducted based on SSR analysis and GMO detection using a total of 13 SSR markers and 2 specific primers, respectively. The SSR samples consisted of 4 samples of sticky rice, 3 samples of brown rice, 8 samples of white rice, and 2 samples of corn, while GMO detection was conducted on 3 samples of corn flour. The aims of this study were to determine the genetic validity between the tested samples compared to the local control rice based on SSR analysis, to confirm the similarity of corn samples based on SSR analysis, and to identify the GMO content of corn flour sample based on qualitative PCR. The results of SSR analysis on sticky rice, brown rice, and white rice showed that their DNA band patterns were different from the local controls. This result was also supported by NTSys analysis. The results of SSR analysis on corn samples indicated that there were different DNA banding pattern between the control and sample corn based on six SSR primers. The results of GMO detection showed that three samples of corn flour were negative. Agroforensic can be performed using SSR method based on PCR technique.

## INTRODUCTION

Forensic identification is one technology to identify specific objects from the trace the evidence in one case of crime. At the first case, forensic identification is related to the disclosure of crimes and in the search for kinship [1]. Identification of DNA fingerprinting in forensics can be presented as DNA chain profile containing repetitive and varied sequences among the investigated individuals [2]. In agriculture or bioscience, this identification will be called agroforensic. Agroforensic will identify and solve the problem when one case such as genetic diversity of crops should be declared, and in other situation need some confirmation like as detection of genetic modified organism (GMO) on food product or agrobusiness.

The discovery of polymerase chain reaction (PCR) by Mullis in 1983 is widely opened in forensic DNA analysis [3], especially related to genetic, phylogenetic, and variations between examined individuals. Together, Sir Alex Jeffreys introduced in the field of forensic genetics this technique by studying a set of DNA fragments that proved to have unique characteristics, which were nonrecurring and inherent for each individual, the only exception being monozygotic twins [4]. The PCR technique opened new ways in DNA analysis in the forensic genetics. The DNA is extracted to be subjected to amplification reactions. The method is an enzymatic process by which regions of the DNA are replicated, generating about one billion ( $10^9$ ) copies.

In agriculture crops, some of the technologies were used in genetic analysis based on PCR technology, such as restriction fragment length polymorphism (RFLP), amplified fragment length polymorphism (AFLP), random amplified polymorphic DNA (RAPD), inter-simple sequence repeats (ISSR), single nucleotide polymorphism

(SNP), simple sequence repeats (SSR), and others [5]. Microsatellites or SSR have been widely used in many crops, due to their abundance, high degree of polymorphism, locus specificity, reproducibility, and most of all, they are a codominant in nature. These make microsatellites an attractive option for genetic studies in date palms [5–7].

Genetic engineering of plants has developed rapidly as technology advances in molecular biology. This discovery led to promote DNA restriction and cloning in a vector. The next process is the development of transformation plants. After all, the confirmation of the present of DNA should be done using simple method such as PCR technique. Until recently, GM crops still debatable regarding to the perspective of people. In Indonesia, handling GM crops needs to use a precautionary approach to realize environmental, food and/or feed safety based on valid scientific methods and take into account religious, ethical, sociocultural, and aesthetic principles [8]. Along the benefits of GM crops, human health risk, biodiversity, increase of insect resistance, and herbicide tolerance, are the most potential concerns of these food materials. In addition, the human health risk of inserted gene into food materials in general has not been systematically shown in the literature. Thus, their detection and labelling is required for increasing the consumer awareness. Furthermore, due to growing the number of unauthorized GM rice varieties in the market and ethical issues about providing informed choice to the consumer, development of screening methods, and monitoring programs seems to be essential in this scope. DNA-based PCR is the accurate and most widely used method for GMO testing.

Introduction of GM crops and their products in the markets required to be monitored and need to know the presence and type of GM elements. There are three main types of DNA based GMO testing methods, i.e. qualitative PCR, southern blot, and DNA microarray. Qualitative analysis comprises of specific detection of target DNA sequence in test samples. Qualitative results clearly validate the presence or absence of GM elements under study, comparative to suitable controls and within the detection limits of analytical technique used and test portion analyzed [9]. PCR has higher specificity to acquire reliable results [10]. On GMO detection, *Cauliflower mosaic virus* (CaMV) 35S promoter (P-35S) and *Agrobacterium tumefaciens* nopaline synthase terminator (T-nos) DNA sequence elements for the detection of genetically engineered (GE) crop plant material were showed the widespread use for detection GM crop plant material. Almost forty-four testing laboratories around the world have been surveyed and used testing using two elements in qualitative and quantitative PCR methods [11]. GMO detection were used PCR technique to identify CaMV P-35S and *A. tumefaciens* T-nos that are the most common transgenic elements that can be targeted for GMO screening. These genetic elements are frequently used to indicate whether the analyzed sample contains GM ingredient or not [12]. The aims of this study were to determine the genetic validity between the tested samples of sticky rice, brown rice, and white rice compared to the local control rice based on SSR analysis, to confirm the similarity of corn samples based on SSR analysis, and to identify the GMO content of corn flour sample based on qualitative PCR.

## MATERIALS AND METHODS

### Materials

Two samples of sticky rice (BP1 and BP2), two samples of brown rice (BP7 and BP9), and five samples of white rice (BP3, BP4, BP5, BP6, and BP8) were collected by criminal division of a police office in Northern Jakarta for SSR analysis. KK, KM, and KB samples were used as local controls of sticky rice, brown rice, and white rice, respectively. Sticky rice Matahari (KtM), white rice Ciherang (CH), and white rice Super Ramos (SR) samples were also used in this study.

Sample of corn B (BISI 18 non-Master) was collected by criminal division of a police office in Eastern Java for SSR analysis. BISI 18 Master was used as control of corn (A).

Three samples of corn flour (B1, B2, and B3) were collected from Surya Grain Indonesia for GMO detection. pCambia1301 plasmid was used as positive control for GM material (transgenic element), while H<sub>2</sub>O was used as negative control.

### DNA Isolation and PCR Analysis

DNA isolation was performed based on the CTAB method by Doyle and Doyle [13] and used for PCR analysis. PCR were performed using thirteen SSR primers (RM23501, RM16253, RM12366, p-bnlg 109, p-bnlg 244, p-umc 1265, p-umc 2039, p-umc 2146, p-umc 2225, p-umc 2279, p-umc 2281, p-umc 2325, and p-umc 2377) to detect polymorphism among rice and corn samples following the method of William *et al.* [14]. Meanwhile, PCR

using primers for P-35S and T-nos fragments were performed to detect possible GMO content in the samples following the method of Listanto *et al.* [15, 16].

Each PCR reaction of SSR analysis contained 1× MyTaq mix, 5 μM forward and reverse primers each, and H<sub>2</sub>O up to 15 μl in total volume. The PCR reaction was started with pre-denaturation at 95°C for 4 min, followed by 35 cycles of denaturation at 95°C for 15 sec, 55°C for 15 sec, and 72°C for 20 sec, and concluded at 72°C for 5 min. The results of PCR amplification of sticky rice, brown rice, and white rice samples were separated in 8% acrylamide gel, while corn flour samples were separated in 2% agarose gel. All of them were visualized using ChemiDoc imaging system.

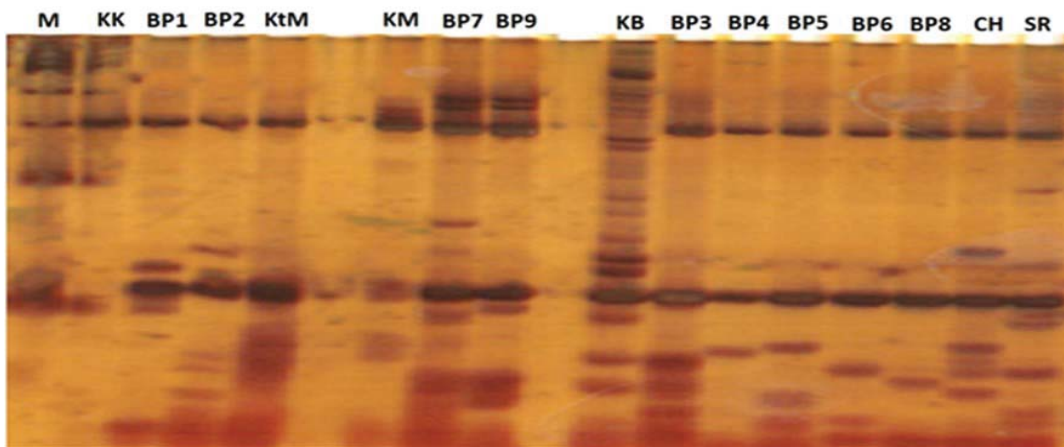
Each PCR reaction of GMO detection contained KAPA Plant PCR Buffer 1× (12.5 μl of KAPA Plant PCR Buffer 2× contains MgCl<sub>2</sub> and dNTPs), KAPA3G Plant DNA Polymerase 0.75 U, 0,3 μM forward and reverse primers each, 100 ng DNA, and H<sub>2</sub>O up to 25 μl in total volume [17]. The PCR reaction was started with pre-denaturation at 95°C for 3 min, followed by 5 cycles of denaturation at 95°C for 15 sec, 60°C for 15 sec, and 72°C for 1 min, followed by 30 cycles of denaturation at 95°C for 15 sec, 55°C for 15 sec, and 72°C for 1 min, and concluded at 72°C for 5 min. The results of PCR amplification of GMO detection were separated in 1.5% agarose gel and visualized using ChemiDoc imaging system.

### NTSys Analysis

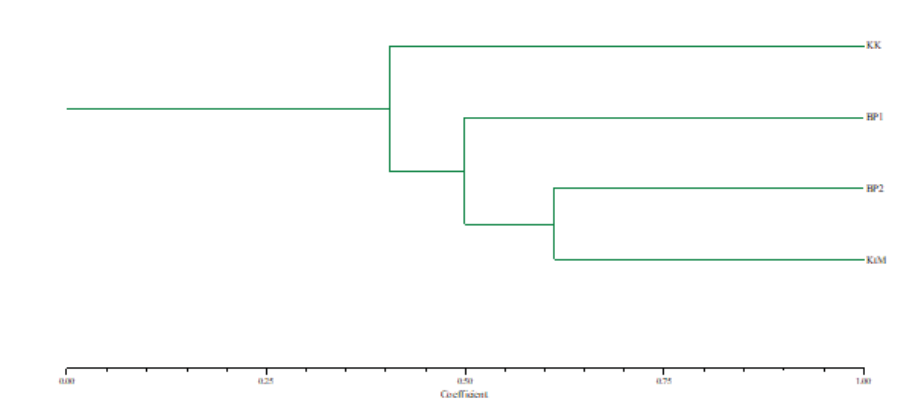
NTSys analysis used the method of Jamshidi [18]. Data entry from agarose gel reads as 0 or 1 to represent the absence or presence of a DNA band, respectively. Clustering analysis included dendrograms drawing procedure such as phylogenetic trees using UPGMA and NTSYS-pc ver. 2.02e program.

## RESULTS AND DISCUSSIONS

Based on the analysis of PCR using SSR primer of RM23501 (Fig. 1), polymorphic bands were produced on all rice samples including the control plants. Further use of additional two different primers (RM16253 and RM12366) on the sticky rice samples showed that the local control of sticky rice (KK) was not in one group with the sticky rice samples of BP1, BP2, and KtM (Fig. 2). They had a similarity level of less than 50% with the local sticky rice according to NTSys analysis. This indicates that all the sticky rice samples were different from the local sticky rice.

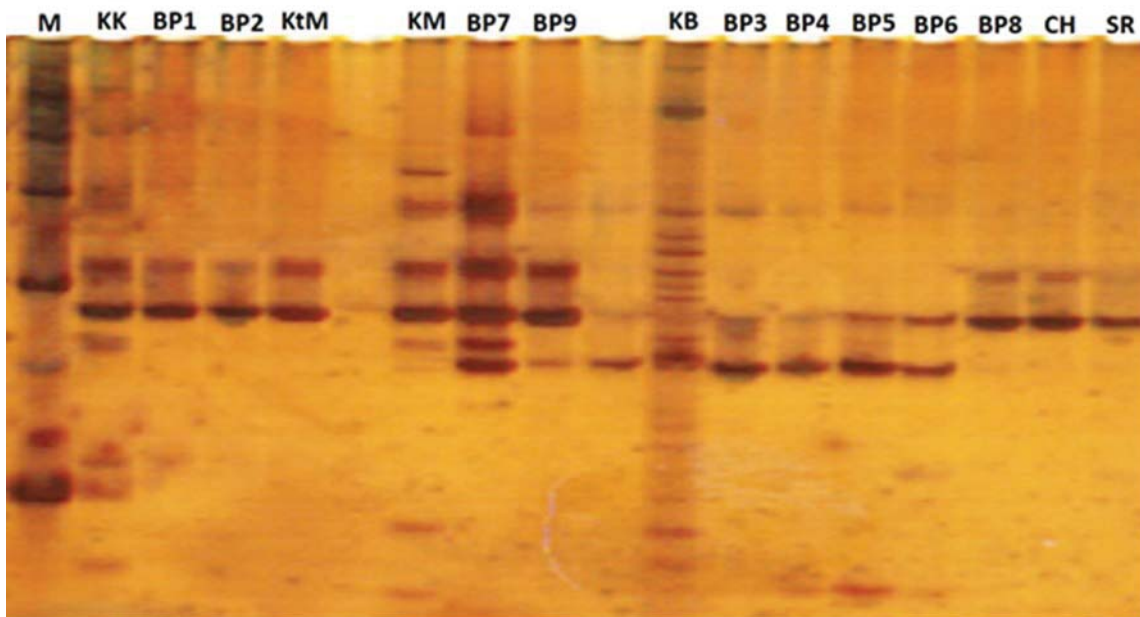


**FIGURE 1.** The results of 8% acrylamide gel electrophoresis based on the PCR product using SSR primer RM23501. M = 100 bp DNA ladder; KK = local control of sticky rice; BP1 and BP2 = sticky rice samples; KtM = sticky rice Matahari; KM = local control of brown rice; BP7 and BP9 = brown rice samples; KB = local control of white rice; BP3, BP4, BP5, BP6, and BP8 = white rice samples; CH = white rice Cihorang; SR = white rice Super Ramos.



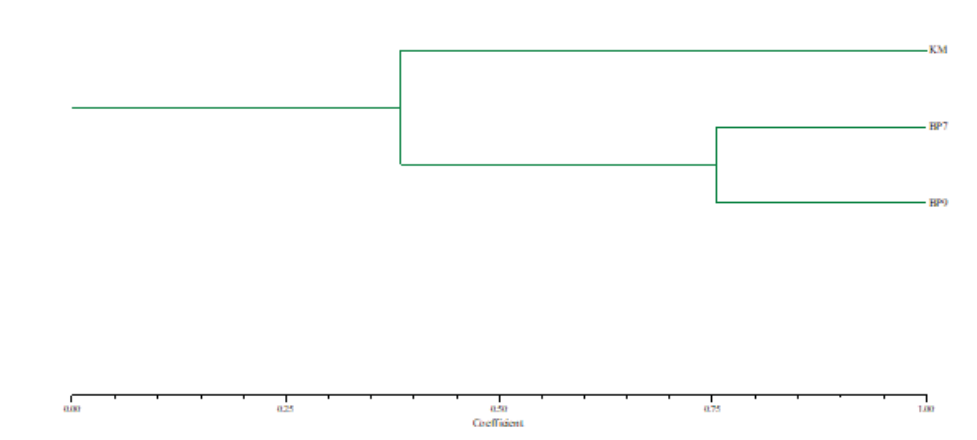
**FIGURE 2.** Dendrogram of sticky rice samples based on SSR analysis using three different primers (RM23501, RM16253, and RM12366). KK = local control of sticky rice, BP1 and BP2 = sticky rice samples, KtM = sticky rice Matahari.

SSR analysis using primer of RM16253 (Fig. 3) showed that the sticky rice samples (BP1, BP2, and KtM) produced same DNA bands, but they had differences of the DNA banding patterns from the local sticky rice (KK). The samples of the brown rice (KM, BP7, and BP9) showed differences of the DNA banding patterns. Meanwhile, the white rice samples of BP3, BP4, BP5, and BP6 had the similarity of the DNA banding patterns. The white rice samples of BP8, CH, and SR have a similar DNA banding pattern. However, the two groups had different banding patterns.



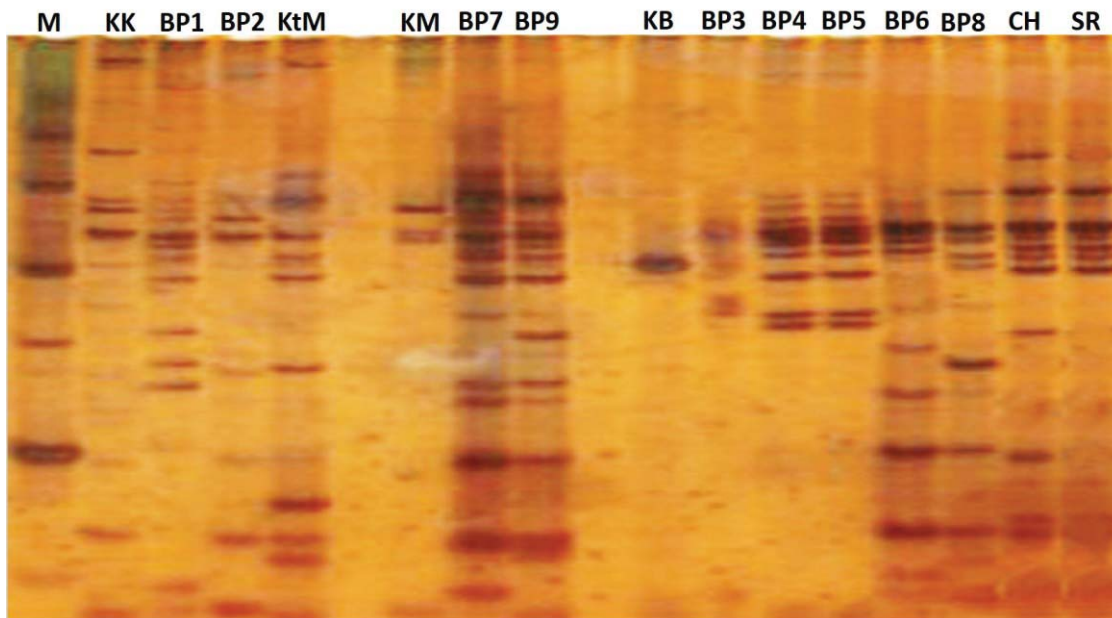
**FIGURE 3.** The results of 8% acrylamide gel electrophoresis based on the PCR product using SSR primer RM16253. M = 100 bp DNA ladder; KK = local control of sticky rice; BP1 and BP2 = sticky rice samples; KtM = sticky rice Matahari; KM = local control of brown rice; BP7 and BP9 = brown rice samples; KB = local control of white rice; BP3, BP4, BP5, BP6, and BP8 = white rice samples; CH = white rice Ciherang; SR = white rice Super Ramos.

The result of SSR analysis using three different primers (RM23501, RM16253, and RM12366) on the brown rice samples showed that the local brown rice (KM) was not in one group with the sample of brown rice of BP7 and BP9 (Fig. 4). Based on dendrogram of NTSys analysis, they only had a similarity level of less than 50%.



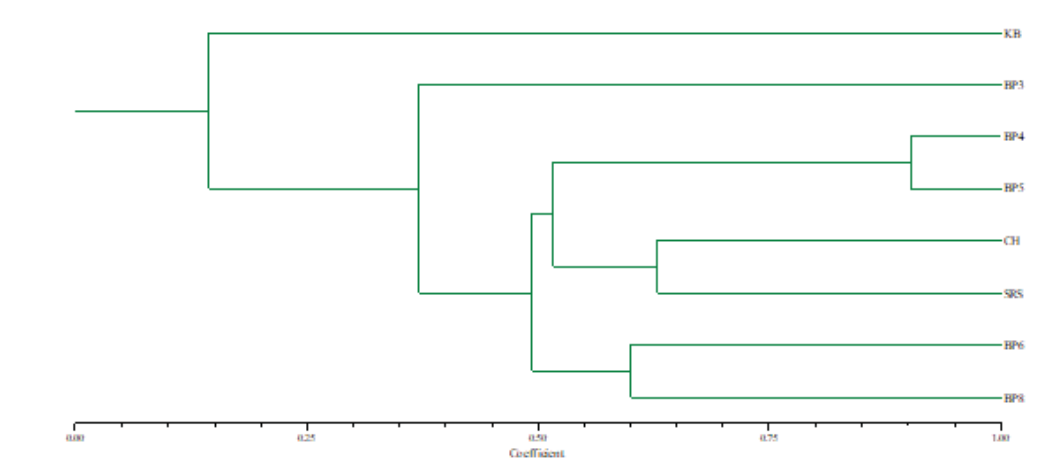
**FIGURE 4.** Dendrogram of brown rice samples based on SSR analysis using three different primers (RM23501, RM16253, and RM12366). KM = local control of brown rice, BP7 and BP9 = brown rice samples.

The PCR results using SSR primer of RM12366 (Fig. 5) showed that all of the amplified rice samples by PCR were polymorphic. However, BP4 and BP5 samples produced monomorphic bands.



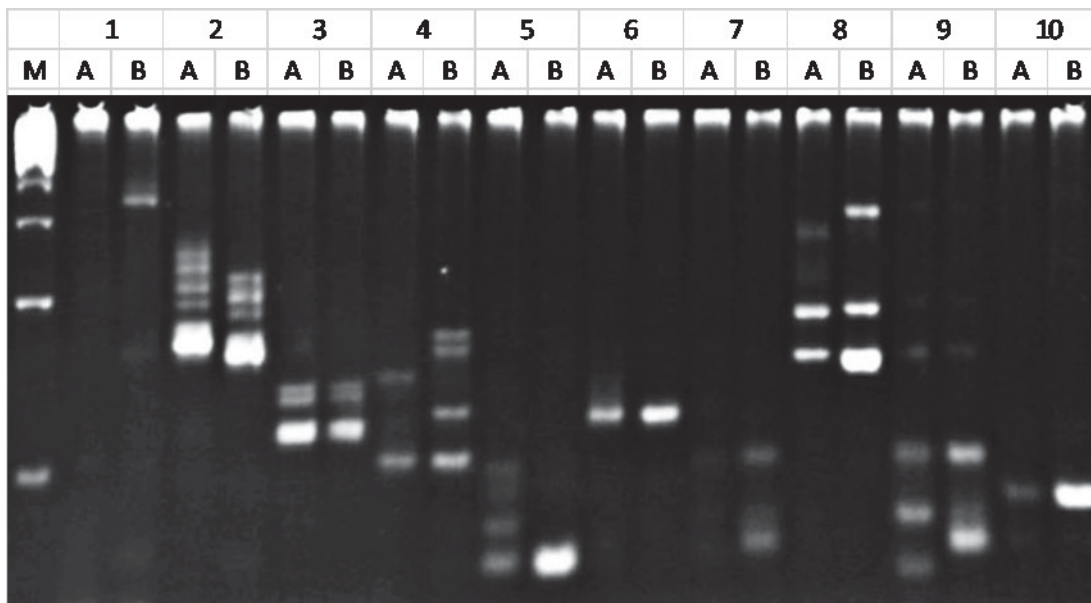
**FIGURE 5.** The results of 8% acrylamide gel electrophoresis based on the PCR product using SSR primer RM12366. M = 100 bp DNA ladder; KK = local control of sticky rice; BP1 and BP2 = sticky rice samples; KtM = sticky rice Matahari; KM = local control of brown rice; BP7 and BP9 = brown rice samples; KB = local control of white rice; BP3, BP4, BP5, BP6, and BP8 = white rice samples; CH = white rice Ciherang; SR = white rice Super Ramos.

The results of SSR analysis using three different primers (RM23501, RM16253, and RM12366) were presented as a dendrogram on Fig. 6. The local control of white rice (KB) was not in one group with the samples of white rice (BP3, BP4, BP5, BP6, BP8, CH, and SR). The control only had a similarity level of less than 25% with the samples. This analysis showed that all the white rice samples were different from the control (KB). In addition, the genetic distance between the BP4 and BP5 samples analyzed using these three primers was similar to that analyzed using one primer (RM12366).



**FIGURE 6.** Dendrogram of white rice samples based on SSR analysis using three different primers (RM23501, RM16253, and RM12366). KB = local control of white rice; BP3, BP4, BP5, BP6, and BP8 = white rice samples; CH = white rice Ciherang; SR = white rice Super Ramos.

The PCR results of corn samples using 10 SSR primers were shown in Fig. 7. The result of PCR amplification of the corn sample B (BISI 18 non-Master) were polymorphic compared to the control A (BISI 18 Master) when using primers 1, 2, 4, 5, 8, and 9 (p-bnlg 109, p-bnlg 244, p-umc 2039, p-umc 2146, p-umc 2281, and p-umc 2325, respectively). Meanwhile, the PCR results using primers 3, 6, and 10 (p-umc 1265, p-umc 2225, and p-umc 2377, respectively) showed that the sample and the control of corn had a similar of DNA binding pattern. Primer 7 (p-umc 2279) did not produce clear DNA binding patterns.

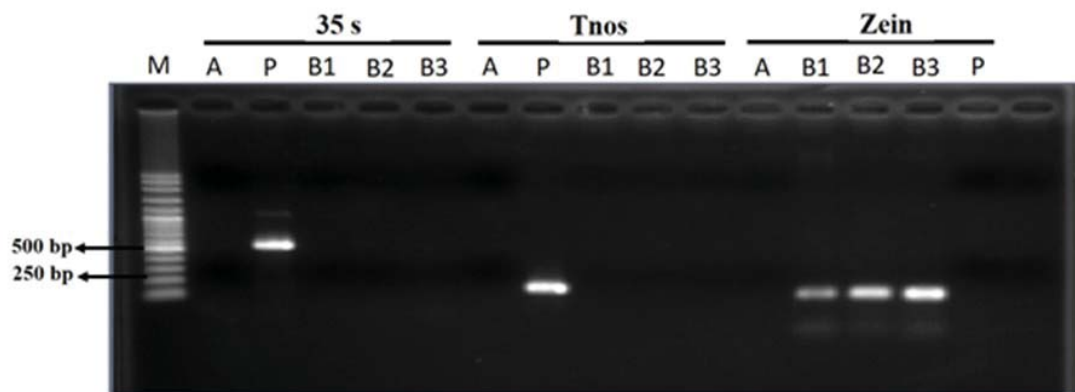


**FIGURE 7.** The results of 2% agarose gel electrophoresis based on the PCR product using 10 SSR primers. M = 100 bp DNA ladder, A = control of corn (BISI 18 Master), B = sample corn (BISI 18 non-Master), 1 = primer p-bnlg 109, 2 = primer p-bnlg 244, 3 = primer p-umc 1265, 4 = primer p-umc 2039, 5 = primer p-umc 2146, 6 = primer p-umc 2225, 7 = primer p-umc 2279, 8 = primer p-umc 2281, 9 = primer p-umc 2325, 10 = primer p-umc 2377.

The use of molecular markers in the field of agricultural forensic, especially SSR markers, can be beneficial to trace the existence of evolution and phylogeny. The previous identification of a sample is based on changes in

geography and morphology of several organisms, so that the molecular markers can be applied to support phylogeny based on genome sequences which are simple and stable in nature. Molecular markers can also be used to investigate the diversity and superiority of a plant variety, such as SSR used in the investigation of heterotic groups of rice and its patterns. Importantly, molecular markers can be used in the investigation of genetic diversity in a large population of germplasm to group the genetic material. Based on these uses, molecular markers, especially SSR, are capable to determine the truth of an organism genetically or certain genetic grouping [5].

On the GMO detection, identification of the presence of P-35S and T-nos fragments is widely used to detect the GM material (transgenic element) of the tested sample. Based on PCR results on B1, B2, and B3 samples of corn flour using primers for P-35S and T-nos fragments, there was no indication that the three samples contained those fragments (Fig. 8). The use of Zein primer to detect corn samples showed that all samples produced DNA fragment of 250 bp, while pCambia1301 plasmid did not produce it (Fig. 8). This confirmed that the samples used were derived from maize/corn.



**FIGURE 8.** The results of 1.5% agarose gel electrophoresis based on the PCR product of the corn flour samples for GMO detection. M = 100 bp DNA ladder; 35s = primer for P-35S fragment; Tnos = primer for T-nos fragment; Zein = primer for Zein fragment; A = H<sub>2</sub>O, negative control; P = pCambia1301 plasmid, positive control for GM material (transgenic element); B1, B2, and B3 = corn flour samples.

**TABLE 1.** Result of GMO detection of the corn flour samples using primers for P-35S, T-nos, and Zein fragments.

Sample	Fragment of P-35S	Fragment of T-nos	Fragment of Zein
B1	Negative	Negative	Positive
B2	Negative	Negative	Positive
B3	Negative	Negative	Positive
pCambia1301*	Positive	Positive	Negative
H <sub>2</sub> O**	Negative	Negative	Negative

\*Positive control for GM material (transgenic element). \*\*Negative control.

Table 1 summarized the PCR results using primers for P-35S, T-nos, and Zein fragments on the three corn flour samples and two control samples. The PCR based methods were used to detect GMO content because the method is widely applicable. This could be applied for unprocessed and highly processed foods. Two types of PCR could be used to examine the presence of GMO: the traditional PCR and the real-time PCR that is capable not only to detect the GMO content, but also to quantify it [19]. In this study, detection of GM material used screening technique based on the existence of P-35S fragment from CaMV and T-nos fragment from *A. tumefaciens* [20]. This method is the most frequently used method for detecting GM material [12].

Several GMO products contained specific GM material in a product in raw material or their derivatives. The common GM materials on the market was gene fragments for insect resistance and/or herbicide tolerance. The application of PCR method that can be used to detect *cry* gene and *EPSPS* gene has been developed using specific primers to verify the absence or presence of these GM materials for product including raw materials (grains) and

their derivatives is needed. In one study, PCR was used to clarify the absence or presence *cry* gene fragment using *cryIAb* primer and herbicide tolerance element using *EPSPS* primer [21].

## CONCLUSION

The results of SSR analysis using three primers (RM23501, RM16253, and RM12366) on sticky rice, brown rice, and white rice showed that the DNA banding pattern was different from their local controls. SSR analysis on corn flour samples using 10 primers indicated that 6 primers produced polymorphic patterns among the control and the samples of corn. In the GMO detection, three samples of corn flour showed negative PCR when using primers for P-35S and T-nos fragments. In summary, agroforensic can be performed using PCR-based SSR method.

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## REFERENCES

1. R. Dumache, V. Ciocan, C. Muresan and A. Enache, "Molecular genetics and its applications in forensic sciences," in *Forensic Analysis – From Death to Justice* (IntechOpen, London, 2016), pp. 87–96.
2. L. Roewer, *Investig. Genet.* **4(1)**, 1–10 (2013).
3. K. Mullis, F. Faloona, S. Scharf, R. Saiki, G. Horn and H. Erlich, *Biotechnol.* **24**, 17–27 (1992).
4. T. Amom and P. Nongdam, *Int. J. Curr. Res. Rev.* **9(17)**, 1–7 (2017).
5. M. A. Nadeem, M. A. Nawaz, M. Q. Shahid, Y. Doğan, G. Comertpay and M. Yıldız, *Biotechnol. Equip.* **32(2)**, 261–285 (2018).
6. R. Manimekalai, P. Nagarajan and P. M. Kumaran, *Trop. Agric. Res.* **18**, 1–10 (2004).
7. S. A. Al-Faifi, H. M. Migdadi, S. S. Algamdi, M. A. Khan, M. H. Ammar and R. S. Al-Obeed, *Electron J. Biotechnol.* **21**, 18–25 (2016).
8. Presiden Republik Indonesia, *Peraturan Pemerintah Republik Indonesia No. 21* (2005).
9. S. Nazir, M. Z. Iqbal and Sajid-ur-Rahman, *Gene Ed. Technol. Appl.* 1–18 (2019).
10. M. Querci, M. Van Den Bulcke, J. Žel, G. Van Den Eede and H. Broll, *Anal. Bioanal. Chem.* **396(6)**, 1991–2002 (2010).
11. M. J. Holden, M. Levine, T. Scholdberg, R. J. Haynes and G. R. Jenkins, *Anal. Bioanal. Chem.* **396(6)**, 2175–2187 (2010).
12. P. Safaei, E. M. Aghaee, G. J. Khaniki, S. A. K. Afshari and S. Rezaie, *J. Environ. Heal. Sci. Eng.* **17(2)**, 847–851 (2019).
13. J. J. Doyle and J. L. Doyle, *Phytochemical Bull.* **19(1)**, 11–15 (1987).
14. J. G. K. Williams, A. R. Kubelik, K. J. Livak, J. A. Rafalski and S. V. Tingey, *Nucleic Acids Res.* **18(22)**, 6531–6535 (1990).
15. E. Listanto, G. A. Wattimena and N. M. Armini, *J. Hortik.* **19(2)**, 137–147 (2009).
16. E. Listanto, E. I. Riyanti, T. J. Santoso, T. Hadiarto and A. D. Ambarwati, *Indones. J. Agric. Sci.* **16(2)**, 51–58 (2015).
17. Kapa Biosystems, <https://www.n-genetics.com/products/1104/1023/13695.pdf> (2016).
18. S. S. Jamshidi, *Int. Conf. Environ. Comput. Sci.* **19**, 165–169 (2011).
19. Y. A. Khidr, M. M. Arafa, S. M. M. Eldemery and R. M. Elsanhoty, *Egypt J. Genet. Cytol.* **47**, 57–68 (2018).
20. C. Xu, L. Li, W. Jin and Y. Wan, *Int. J. Mol. Sci.* **15(10)**, 18197–18205 (2014).
21. Bahagiawati, Reflinur and T. J. Santoso, *J. AgroBiogen* **11(2)**, 65–72 (2015).