

# The Second International Conference on Genetic Resources and Biotechnology

## Harnessing Technology for Conservation and Sustainable Use of Genetic Resources for Food and Agriculture

Bogor, Indonesia • 24–25 May 2021

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January 2022

THE SECOND INTERNATIONAL CONFERENCE ON GENETIC RESOURCES  
AND BIOTECHNOLOGY: Harnessing Technology for Conservation and Sustainable  
Use of Genetic Resources for Food and Agriculture

# Committees: The Second International Conference on Genetic Resources and Biotechnology

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## **Preface: The Second International Conference on Genetic Resources and Biotechnology**

The Second International Conference on Genetic Resources and Biotechnology, which is the continuation of the first event held in 2018, focuses on topics related to advances in biotechnology to create more opportunities for effective conservation and sustainable utilization of genetic resources for food and agriculture. This year conference's theme is Harnessing Technology for Conservation and Sustainable Use of Genetic Resources for Food and Agriculture. The conference was organized by Indonesian Agency for Agricultural Research and Development (IAARD), Ministry of Agriculture, Indonesia, in collaboration with Indonesian Biotechnology Consortium and held on 24<sup>th</sup>-25<sup>th</sup> of May 2021 virtually due to the pandemic of COVID-19.

The conference aims to share and exchange current scientific information and technological developments on biotechnology and their applications for conservation and sustainable use of genetic, to encourage and promote quality, efficiency, and modernization of management and utilization of genetic resources, and to facilitate national and international collaboration among participants. There are five scopes discussed in this conference. They are effective management of conservation and sustainable use of genetic resources for food and agriculture, application of genomics and molecular markers for genetic resource conservation and crop adaptation to climate change, application of innovative crop improvement techniques for conservation and sustainable use of plant genetic resources for food and agriculture, plant cell and tissue culture for conservation and effective utilization of genetic resources, and the use of microbial genetic resources as biological control agents of agricultural pests and diseases, and for soil bioremediation.

Five speakers from the United States of America, Japan, India and Indonesia were invited to discuss about their expertise and knowledge on relevant subjects in the plenary sessions. This conference was attended by more than 100 participants including 75 presenters and 44 listeners worldwide. They came from diverse governmental, private, or academic institutions and also scientific communities. The presented materials have undergone peer review processes and only qualified papers were selected. Furthermore, all papers were subjected to double blind peer-review and expected to meet the scientific criteria of significance and academic excellence to be published in a conference proceedings indexed in a well-known, reputable service.

We would like to express our sincere gratitude to our speakers, presenters and all participants for their contributions in this conference. We would also like to express our appreciation for the generosity of our sponsors that support this conference: PT CropLife, PT ITS Science Indonesia, PT Fajar Mas Murni and PT Prima Instrument Analitika. Lastly, special thanks to all committee members for their exceptional work and contributions in the conference and publication.

Chair of Organizing Committee

Dr. Toto Hadiarto

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**Abstract.** Japansche citroen (JC) is popular citrus rootstock in Indonesia due to well-adapted to biotic and abiotic stresses. JC is naturally rich of secondary metabolites. Therefore, cells production of JC through tissue culture is potential for continuous metabolites farming in local pharmaceutical industries. The objectives of study were to determine the best initial cells density (ICD) in flask and air-lift bioreactor (ALB) cultures for mass cells proliferation and to determine the limonin content in cells. Callus was induced from nucellus and habituated callus were collected to initiate cell suspensions. Selected cells were transferred to flask and ALB cultures at the density of 2, 4, and 6 mg/ml for 90 days of culturing. An increasing ICD affected cell growth in flask and ALB cultures. Cells in flask culture grew slower and were produced lower in quantity than in ALB. At 4 mg/ml, growth rate (GR) of cells in flask culture at day 30 reached maximum of 4.12 mg/day, 87-folds lower than in ALB that reached 360 mg/day. Limonin at 120.725 µg/g dried cells (DW) were harvested from ALB within 30 days, while total phenolic was 9.883±0.471 mg GAE/g DW and total flavonoid was 0.144±0.011 mg QE/g DW. This study confirmed the advantages of cell culture as a perpetual source for production of biomass content metabolites that amenable lower operational cost compared to natural production for local industries.

## INTRODUCTION

Citrus is one of the most important fruit in the world for human health due to their active properties. Citrus fruit possess a range of phenolic compounds, alkaloids, flavonoids, terpenes, limonoids, phytonutrients and vitamins C, carotenoids, and essential oils [1]. Limonoid is one of the important compound in citrus. It is a family of polycyclic secondary metabolites [2] that are commonly found in seeds [3] and fruits of pummelos, grapefruits, lemons, sweet oranges, and mandarins [4], peels of ponderosa lemon *Citrus pyriformis* Hassk [5], *C. jambhiri* Lush [6], bergamot (*C. bergamia* Risso) [7], rind and pulp of *C. limon* fruit [8]. About 50 limonoids aglycones and glucosides and even more have been isolated from diverse citrus species, such as *C. aurantium*, *C. limon*, *C. reticulata*, *C. sudachi*, *C. unshiu*, *C. paradisi*, *C. sinensis*, *C. jambhiri*, and *C. pyriformis* [9], and *C. limonia* Osbeck (Japansche citroen/JC) [10]. *C. limonia* cv. Rangpur lime (JC) is popular rootstock [11]. In fact, broaden adaptation of rootstock is driven by its metabolites compound [11].

Limonoids has been found largely in Rutaceae and Meliaceae [9]. The name limonoid comes from limonin, the first bitter tetranortriterpenoid isolated from citrus [12]. Research interest on limonoid from citrus is due to its contribution to the bitter taste of citrus fruits, which harms the global citrus fruit and juice industry [13]. However, it's diverse bio-activities evaluated *in vitro* and *in vivo* possess several biological activities, including antitumor, neuroprotective, immunomodulatory, insecticidal, anti-obesity, anti-hyperglycemic activities [13], analgesic [14], anticancer such as liver cancer [15], breast cancer [16], intestinal carcinogenesis [17], colon cancer [18], anti-inflammatory [13], antibacterial [13], antioxidant [6], antiviral [13], antimalarial [9], and larvicidal [19], prevent non-alcoholic fatty liver, cardiovascular diseases, type-2 diabetes mellitus, and metabolic syndromes [20]. As food-

derived phytochemicals, limonoids from citrus confirmed to have well tolerated by non-cancerous cell lines [2], animal models [21], and human body [22]. Therefore, limonoids present potential applications in food and pharmaceutical industries, functional food additives, pesticides, and feed additives [13]. Recently, molecular docking studies of phytochemicals against the therapeutic protein targets of SARS-CoV-2 supported the effective binding affinity with limonin, a triterpenoid found in citrus [23]. The highest level of genomic similarity between SARS-CoV and SARS-CoV-2 [24], and the effectiveness of triterpenoids against SARS-CoV prompted us to search potential phytochemicals from limonoids and triterpenoids [25].

Production of agricultural based product that safe to environment, biodiversity, and quality of agricultural crops are referred to sustainable crop production [26]. Tissue culture of plant (PTC) offers remarkable prospects to produce desirable plant structures, such as cells [27], shoots [28], root [29], and a variety of secondary metabolites inside the cells in large quantities [30] for drugs, cosmetics, and food. Extracted material derived from plant cell and tissue cultures were extensively used as component of 50 products and brought a new wave in PTC technology [27].

The PTC have been utilized to produce seedling of *C. reticulata* Blanco cv. Madu [31], *Oncidium* ‘Sugar Sweet’ [32], *Musa acuminata* cv. Berangan [33], and *Vitis vinifera* L. [34] using air-lift bioreactor (ALB) culture. The similar method of PTC techniques seem as alternatives method for producing secondary metabolites [35] when natural supply is inadequate or chemical synthesis is impossible [36].

Method of production by PTC has some advantages over natural production and even endorsed by Food and Agriculture Organization [37], i.e. easy and scalable production of secondary metabolites [38], rapidity of production [30], independent of season [27], fully controllable room during process [39], produce chemicals in environmentally safely manner [36], uniformity and standardization of the desired compounds [40], and independent of climatic and/or any various environmental factors [41].

Large-scale liquid cultures have been used for micropropagation through organogenesis [28] or somatic embryogenesis pathways [31]. Flask culture and bioreactor have been employed mainly for cell suspension cultures and secondary metabolites production [42] such as in *Arnebia* sp. [43]. Cell suspension culture is most amenable to large-scale production of cell in term of rapid cell proliferation and large number [44]. The objectives of the present study were to determine the best initial cells density (ICD) in flask and ALB cultures for mass cells proliferation and to determine the limonin content in cells.

## MATERIALS AND METHODS

The studies were conducted in Tissue Culture Laboratory of Indonesian Citrus and Subtropical Fruits Research Institute (ICSFRI or Balitjestro). The pro-embryonic masses were initiated in 2015 and cell proliferations in flask and bioreactor cultures were conducted in 2016. Citrus limonin and metabolites were analyzed from cells derived optimized protocols [10, 45].

### Culture Establishment

Young fruits were collected from mother plant of *C. limonia* cv. JC planted in the experimental field in ICSFRI. Fruits were then washed under running tap water and transferred to laminar air-flow cabinet (LAFB). A sequential sterilization process was subjected to the explants for cleaning the surface following as described previously [31]. Selected normal young seeds were planted on MS basal medium (Duchefa M220) supplemented with 0.5 g/l malt extract (ME), 30 g/l sucrose, 13% (w/v) agar, and 1 mg/l BAP to induce embryogenic calli. The pH of the medium was adjusted to 5.7±0.1 with 0.5 M NaOH before autoclaving at 121°C and 15 psi (103 kPa) for 15 min. The embryogenic calli were sub-cultured biweekly on fresh semisolid medium for selecting homogenous cell growth and habituating pro-embryonic masses (PEM). After 10 subcultures, the PEM were then transferred to liquid culture for another cell adaptation. Cell suspension initiation was conducted as Agisimanto *et al.* [46]. Cultures were sub-cultured biweekly in flasks by removing liquid media and addition of 25 ml fresh liquid medium that similar to medium initiation without agar.

### Flask and Air-Lift Bioreactor Cultures

Experiments of flask culture were conducted to evaluate kinetic growth of cells. Cells at 2.0, 4.0, and 6.0 mg/ml were inoculated to 25 ml liquid MS medium supplemented with 0.5 mg/l BAP and 500 mg/l ME. The quantities of

explants were equal to 50, 100, and 150 mg PEM per 25 ml medium. The cultures were mechanically agitated on a rotary shaker at 100 rpm at 25°C. The liquid media were refreshed biweekly for six times. ALB cultures were conducted after establishing suitable growth of cells in flask cultures for cell proliferation. Experiments on ALBs to verify the best condition for scaling-up were conducted. Explants at similar density to that in flask cultures were inoculated into 3 l ALBs containing 1 l MS basic medium supplemented with 0.5 mg/l BAP and 500 mg/l ME. The quantities of explants were equal to 2, 4, and 6 mg/ml in 2 l medium. The cultures were supplemented with continuous aeration at 3 vvm. Air was pumped into the ALB via a 0.22 µm filter. A flowmeter was placed between the air filter and pump on each culture to control the rate of aeration. The liquid media were refreshed monthly during 3 months of culture. After 30 days of initiation, liquid media were increased to 2 l for maintaining cells suspension. All the cultures were incubated at 23±2°C under continuous illumination with cool white fluorescent lamps with light intensity at 35 µmol/m<sup>2</sup>/s.

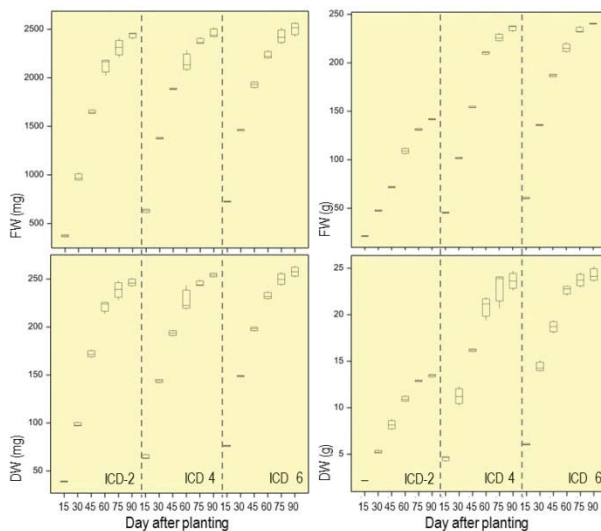
## Experimental Setup, Data Collection, and Statistical Analysis

In order to compare the kinetic growth of JC in flask and ALB cultures, two separate experiments were carried out under a completely randomized design with three replications per treatment. About 162 flasks containing 25 ml medium were inoculated with cells at three level of ICD (2, 4, and 6 mg/ml), repeat 3 times, 54 units of ALB containing 1,000 ml medium were also inoculated at similar ICD as flask cultures mentioned above. After 15, 30, 45, 60, 75, and 90 days of culture, three samples of flask and an ALB culture were collected and dried on filter paper for 2 h biweekly to measure the kinetic growth of cells, expressed as fresh cell (FW) each treatment and dried cells (DW) after kept in a dryer at 50–60°C for at least 6 h. The biweekly growth rate (GR) of cells was calculated as final weight ( $W_f$ )–initial weight ( $W_i$ ) per time (t).

Data were analyzed individually per set of treatment by analysis of variance, and multiple comparison analysis was made using least significant differences (LSD) using Genstat 19<sup>th</sup> edition. Probabilities less to or equal to 0.05 were considered significant.

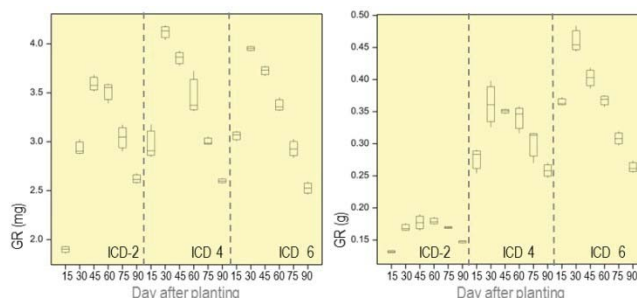
## RESULTS AND DISCUSSIONS

Growth and cell proliferation in liquid medium were significantly affected by ICD. Pattern of cell growth represented by FW and DW in flask and ALB cultures increased exponentially following sigmoid curves (Fig. 1). The patterns of sigmoid curves were specific and significantly different in every treatment of ICD, both in flask and ALB cultures. In this study, inoculation of cells at 2, 4, and 6 mg/ml into flask cultures that were refreshed every 15 days, increased the biomasses significantly until day 75 of cultures.



**FIGURE 1.** Fresh cells weight (FW) and dried weight (DW) of *Citrus limonia* cv. Japanche citroen at ICD-2 (2 mg/ml), ICD-4 (4 mg/ml), and ICD-6 (6 mg/ml) under flask (left) and air-lift bioreactor (ALB) cultures (right) during 90 days of cell suspension culture.

The FW, DW, and GR of both cultures increased linearly at day 15 to 45 after initiation. However, the cells in ALB cultures increased faster in early cycle of culture compared to flask cultures. The GRs of ALB cultures were the highest at day 30 of cultures then reduced linearly (Fig. 2), while cells in flask cultures increased up to 45 days of culture, then the GR reduced by the addition of culture period (Fig. 2). After 75 days of culture, the cell number increased stationary and not significantly based on multiple comparison analysis.



**FIGURE 2.** Growth rate (GR) of *Citrus limonia* cv. Japanche citroen cells from different at ICD-2 (2 mg/ml), ICD-4 (4 mg/ml), and ICD-6 (6 mg/ml) under flask (left) and air-lift bioreactor condition (right) during 90 days of cell suspension culture.

The total amount of FW and DW of cells inoculated with 2 mg/ml cells increased significantly until 45 days compared to 4 and 6 mg/ml cells, while FW and DW of cells inoculated in flask culture with ICD of 4 and 6 increased only up to 15 days of culture. After 15 days of culture, the cells grew similarly (Tables 1 and 2). Lower concentration of ICD in starting culture produced lower FW and DW at day 90 of culture. However, the GR of 2 mg/ml was still higher until the end of observation (Table 3). The addition of ICD from 2 mg/ml to 4 mg/ml increased the GR from 3.59 mg/day to 3.86 mg/day at 45 day after initiation. Meanwhile, a further increase of ICD from 2 mg/ml to 6 mg/ml of ICD reduced yields by 3.73 mg/day compared to 4 mg/ml of ICD. Based on the DW data at day 45 of culture, cultures inoculated with 2.0 mg ICD/ml multiplied 34-folds while at 4 or 6 mg/ml, the rate of cell multiplication were 19-folds and 13-folds respectively (Table 2).

In ALB cultures, FW, DW, and GR were significantly affected by ICD. In the present study, the total amount of FW and DW of cells biomasses harvested from ALB cultures and GR increased with increasing cells density. The FW at day 30 increased by 25-folds and 22-folds when cell density was increased from 2 to 4 and 6 mg/ml, respectively, whereas at 2 mg/ml of cell density, FW increased by only 23 folds (Table 1). The DW at day 30 of cultures increased similarly to FW (Fig. 1). The GR of cultures inoculated with cells at 2, 4, and 6 mg/ml reached maximum value, i.e. 170, 360, and 400 mg/day, respectively, which is 57, 87, and 116 times higher than those of flask cultures at day 30 (Fig. 2, Table 3). The rapid growth of cell biomasses in ALB cultures compared to flask cultures was significantly different even though the same amount of cells were administered.

Cell growth in ALB was enhanced rapidly with an increase of ICD from 2 to 6 mg/ml. At low ICD (2 mg/ml), the final yield was lower, i.e. 141.7 g at day 90 of culture, compared to 235.8 and 240.4 g of fresh cells at ICD of 4 and 6 mg/ml, respectively. On the contrary, the increase in inoculum size in flask culture was not accompanied by higher cell growth rate. It is also shown that DW at initial density of 4 mg/ml grew faster in the first period of 30 days after inoculation (DAI), while at density 2 and 6 mg/ml grew slower. Even at the lower initial cell inoculums (4 mg/ml) the cells grew faster in the early stage of growth. Nevertheless, the total amount of cells produced in the 6 mg/ml inoculum culture was still higher than that in the other treatments (Table 2). Based on GR parameter, ALB cultures inoculated with 4 mg/ml of ICD reached maximum GR at day 30 of culture, at the rate of 0.36 g/day, while at the same level of ICD, flask culture reached maximum GR of 4.12 mg/day. At ICD of 4 mg/ml, cell growth followed a linier curve during a period of culture at day 15 to 45. This type of growth curve was important to get faster cell growth to reach more than 150 g cells. At ICD of 6 mg/ml, the cells grew slower. This study indicated that lower ICD stimulated faster cell growth in the early stages of culture. However, the total amount of cells produced at the higher inoculum was still higher than at the lower inoculum.

**TABLE 1.** Inoculum cell density effect on the fresh weight of cells of *Citrus limonia* cv. Japansche citroen in flask and air-lift bioreactor (ALB) cultures.

| Inoculum cell density (mg/ml) | Days after planting*          |         |         |          |          |         |
|-------------------------------|-------------------------------|---------|---------|----------|----------|---------|
|                               | 15                            | 30      | 45      | 60       | 75       | 90      |
| <b>Flask</b>                  | <b>Cell fresh weight (mg)</b> |         |         |          |          |         |
| 2                             | 273 a                         | 977 a   | 1,650 a | 2,125 a  | 2,311 a  | 2,438 a |
| 4                             | 630 b                         | 1378 b  | 1,886 b | 2,162 ab | 2,378 ab | 2,461 a |
| 6                             | 726 c                         | 1462 b  | 1,929 b | 2,235 bc | 2,427 bc | 2,504 b |
| P-value                       | <0.01                         |         |         |          |          |         |
| CV (%)                        | 3.10                          |         |         |          |          |         |
| <b>ALB</b>                    | <b>Cell fresh weight (g)</b>  |         |         |          |          |         |
| 2                             | 21.4 a                        | 47.7 a  | 71.8 a  | 109.1 a  | 131.3 a  | 141.7 a |
| 4                             | 45.5 b                        | 101.8 b | 154.5 b | 210.1 b  | 226.0 b  | 235.8 b |
| 6                             | 60.6 c                        | 135.8 c | 187.0 c | 215.2 c  | 234.0 c  | 240.4 c |
| P-value                       | <0.001                        |         |         |          |          |         |
| CV (%)                        | 1.60                          |         |         |          |          |         |

\*Means in the same column followed by the same letter are not significantly different at  $P \leq 0.05$  according to the Duncan's Multiple Range Test.

**TABLE 2.** Inoculum cell density effect on the dry weight of *Citrus limonia* cv. Japansche citroen in flask and air-lift bioreactor (ALB) cultures.

| Inoculum cell density (mg/ml) | Days after planting*        |         |         |          |          |          |
|-------------------------------|-----------------------------|---------|---------|----------|----------|----------|
|                               | 15                          | 30      | 45      | 60       | 75       | 90       |
| <b>Flask</b>                  | <b>Cell dry weight (mg)</b> |         |         |          |          |          |
| 2                             | 39 a                        | 98.5 a  | 172.1 a | 221.3 a  | 238.6 a  | 246.4 a  |
| 4                             | 54.7 b                      | 143.7 b | 193.6 b | 228.3 ab | 245.4 ab | 254.2 ab |
| 6                             | 76.1 c                      | 148.8 b | 197.9 b | 232.7 bc | 249.9 bc | 257.6 c  |
| P-value                       | <0.001                      |         |         |          |          |          |
| CV (%)                        | 3.00                        |         |         |          |          |          |
| <b>ALB</b>                    | <b>Cell dry weight (g)</b>  |         |         |          |          |          |
| 2                             | 2.17 a                      | 5.27 a  | 8.17 a  | 10.96 a  | 12.90 a  | 13.43 a  |
| 4                             | 4.54 b                      | 11.24 b | 16.18 b | 20.82 b  | 22.89 b  | 23.64 b  |
| 6                             | 6.08 c                      | 14.42 c | 18.71 c | 22.65 c  | 23.73 bc | 24.30 c  |
| P-value                       | <0.001                      |         |         |          |          |          |
| CV (%)                        | 5.30                        |         |         |          |          |          |

\*Means in the same column followed by the same letter are not significantly different at  $P \leq 0.05$  according to the Duncan's Multiple Range Test.

**TABLE 3.** Inoculum cell density effect on the growth rate of *Citrus limonia* cv. Japansche citroen on flask and air-lift bioreactor (ALB) cultures.

| Inoculum cell density (mg/ml) | Days after planting*           |        |         |         |        |        |
|-------------------------------|--------------------------------|--------|---------|---------|--------|--------|
|                               | 15                             | 30     | 45      | 60      | 75     | 90     |
| <b>Flask</b>                  | <b>Cell grow rate (mg/day)</b> |        |         |         |        |        |
| 2                             | 1.90 a                         | 2.94 a | 3.59 a  | 3.51 c  | 3.04 b | 2.62 a |
| 4                             | 2.98 c                         | 4.12 c | 3.86 bc | 3.47 ab | 3.01 b | 2.60 a |
| 6                             | 3.06 c                         | 3.95 b | 3.73 b  | 3.37 a  | 2.93 a | 2.53 a |
| P-value                       | <0.01                          |        |         |         |        |        |
| CV (%)                        | 3                              |        |         |         |        |        |
| <b>ALB</b>                    | <b>Cell grow rate (g/day)</b>  |        |         |         |        |        |
| 2                             | 0.13 a                         | 0.17 a | 0.18 a  | 0.18 a  | 0.17 a | 0.15 a |
| 4                             | 0.28 b                         | 0.36 b | 0.35 b  | 0.34 b  | 0.30 b | 0.26 b |
| 6                             | 0.37 a                         | 0.37 b | 0.40 c  | 0.37 b  | 0.31 b | 0.26 b |
| P-value                       | <0.001                         |        |         |         |        |        |
| CV (%)                        | 5.5                            |        |         |         |        |        |

\*Means in the same column followed by the same letter are not significantly different at  $P \leq 0.05$  according to the Duncan's Multiple Range Test.

At the time of inoculation, a plant cell culture needs a minimum inoculum concentration as the level of nutrient per cell is important for the initial cell division [47]. Cell culture needs a minimum number of cells in the medium at the time of inoculation, which determines the level of nutrition required for each cell for initial cell division. The stimulation of cell growth with increased ICD might result from cell-cell communication among the inoculum cells [48]. The increase in ICD has been shown to achieve higher levels of cell proliferation in other citrus [49] and even more in bioreactor as reported in ginseng [50].

Our study also found that ALB supported rapid and faster cell growth and proliferation and also larger quantity of cells than flask cultures, for several advantages reasons as mentioned earlier [44]. ALB culture provides beneficial for massive production system of healthy organs and the growth of valuable plants throughout the year [51]. In order to obtain the highest yield of production, those factors mentioned above should be determined prior to the application on a bioreactor [52]. The best conditions for cell, organ and/or plant growth and development can be conveniently achieved in a bioreactor by completely regulating several chemical and physical factors, including mixing, gaseous composition, efficient oxygen transfer, pH, and hydrodynamic forces [50].

From the optimized ALB cultures (Fig. 3), the 30 days period of culture produced limonin at 120.725  $\mu\text{g/g DW}$ , while total phenolic was  $9.883 \pm 0.471$  mg GAE/g DW and total flavonoid was  $0.144 \pm 0.011$  mg QE/g DW. These results were lower than the secondary metabolite production from natural leaves that produced limonin at 250.980  $\mu\text{g/g DW}$ , total phenolic at  $9.883 \pm 0.470$  mg GAE/g DW, and total flavonoid at  $8.152 \pm 0.918$  mg QE/g DW.



**FIGURE 3.** Cell proliferation of *Citrus limonia* cv. Japansche citroen in 100 ml flask at 30 days of culture (left) and in 3 l ALB culture (right).

As bio-process optimization and scale-up of plant cell cultures containing secondary metabolites require an understanding of substrate and inoculums requirements for the production of cell mass, our recent finding confirmed the importance of cell density to produce elite cells in short period of time at high quantity. This information is important for commercial purposes in terms of biomasses and/or secondary metabolites production in bioreactor and amenable lower operational cost compared to natural production. This would be also possible for citrus-based material in Indonesia.

## CONCLUSION

Increased cell density affected cell growth in flask and ALB cultures. Flask culture grew slower than ALB culture and produced lower quantity of biomass. The maximum GR in flask culture reached maximum of 4.12 mg/day at day 30 of culture inoculated with 4 mg/ml of ICD. While at the same day, the best GR of cell in ALB culture reached 360 mg/day at cultures inoculated with 4 mg/ml of ICD. ICD at 100 mg per 25 ml medium in flask culture produced 1.378 mg FW cells, 87-folds lower than ALB culture inoculated with 4 g/l medium. From optimized ALB culture, the 30 days period of culture produced limonin of 120.725  $\mu\text{g/g DW}$ , while the total phenolic was  $9.883 \pm 0.471$  mg GAE/g DW and the total flavonoid was  $0.144 \pm 0.011$  mg QE/g DW.

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