

BULETIN *AgroBio*

ISSN 0853-9022

Vol. 4, No. 2, 2001

JURNAL TINJAUAN ILMIAH RISET BIOLOGI DAN BIOTEKNOLOGI PERTANIAN

The Use of Isozymes as Biochemical Markers in Rice Research Buang Abdullah	39
Pembentukan Buah Partenokarpi melalui Rekayasa Genetika Saptowo Jumali Pardal	45
Paradigma Baru Pengendalian Wereng Batang Coklat (<i>Nilaparvata lugens</i> Stål.): Endosimbion sebagai Sasaran Mohammad Iman dan Tri Puji Priyatno	50
Aplikasi Pupuk Hayati dan Pupuk Kimia: Suatu Pendekatan Terpadu R.D.M. Simanungkalit	56
Perbaikan Teknologi Produksi Kacang Tanah di Indonesia Muhammad Sudjadi dan Yati Supriati	62



Balai Penelitian Bioteknologi Tanaman Pangan
Badan Penelitian dan Pengembangan Pertanian

The Development of Insect-resistant Plants through Biotechnology

Sutrisno

Research Institute for Food Crop Biotechnology, Bogor

ABSTRAK

Pembentukan Tanaman Tahan Serangga Hama melalui Bioteknologi. *Sutrisno.* Sejumlah tanaman tahan serangga hama telah ditanam di seluruh dunia sejak beberapa waktu yang lalu. Pembentukan tanaman tahan serangga hama akan terus berlanjut untuk mengantisipasi timbulnya serangga hama biotipe baru dan pergeseran status serangga hama utama. Bioteknologi menawarkan alat ampuh untuk digunakan dalam pembuatan tanaman tahan serangga hama secara komplementer dengan metode konvensional. Alat bioteknologi itu ialah kultur anter, fusi protoplas, kultur embrio, variasi somaklonal, seleksi dibantu markah, dan transformasi genetik. Pilihan tiap alat itu dapat didasarkan pada banyak alasan antara lain efisiensi, kecepatan, kepastian, dan asal gen-gen ketahanan. Tanaman tahan serangga hama dihasilkan dari pendekatan konvensional atau bioteknologi mungkin menyebabkan efek serupa dalam pembentukan serangga hama biotipe baru.

Kata kunci: Tanaman tahan serangga hama, bioteknologi

The development of insect-resistant plants has been started in 1782 since Havens published a paper about a wheat cultivar resistance to the Hessian fly. Since then, numerous insect-resistant plants have been developed by the international and national research center, private sector through conventional method or biotechnology. For example, International Rice Research Institute (IRRI) has developed and released numerous rice variety resistance to brown planthopper, green leafhopper, rice stem borer, and rice gallmidge. Several transgenic insecticidal cultivars obtained through biotechnology approach have also been developed, such as Bt-corn, Bt-cotton, and Bt-rice.

Many insect-resistant plants have been adopted by farmers at all over the world. Rice resistance to the rice brown planthopper and green rice leafhopper has been planted over 20 million hectare in Asia. Wheat resistant to Hessian fly are grown in the mid-western of United States. In 1999, Bt-plants resistance to insect has been planted over 8.9 million hectare mostly in United States (James and Krattiger, 1996). The wide spread use of insect-resistant plants may be due to the fact that those plants are the most

economic, least complicated and environmentally soundest approach for protecting plants from insect damage.

The risks may arise from the use of insect-resistant plants derived from conventional method have been well documented that includes the development of new insect biotypes, such as brown planthopper biotypes and Hessian fly biotypes. However, up to now the development of new insect biotypes that overcoming transgenic insecticidal plants derived from biotechnology approach has not been recorded. Two strategies to delay the development of new insect biotypes have been reported. The first strategy relate to insect-plant resistance development and the second strategy relate to their application in the field in which insect-resistant plants as one of the component of integrated pest management approach.

Controversy in the use of transgenic insecticidal plants derived from biotechnology approach is still continuing in the world, including in Indonesia. This paper was presented with the aim to get better understanding to transgenic insecticidal plants as one of insect-resistant plants.

METHOD TO DEVELOP INSECT-RESISTANT PLANTS

In breeding program for insect-resistant plants, the first step is to identify the parents or donors of resistance. These may be cultivated germplasm, landraces, weed races, wild species, or different species. The breeding method used to develop insect-resistant plants based on the following factors: the source of donor of resistance, the efficiency and certainty of selection of progenies. If the donor of resistance is commercial varieties, they may be improved by pure line or mass selection or hybridized with elite germplasm. If they are landraces, weed races, or wild species, they have to be hybridized with the elite germplasm. If the donor of resistance is different species, biotechnology method may be used, such as genetic transformation or protoplast fusion. If the donors are not available, biotechnology method may be used, such as somaclonal variation for insect resistance. If the efficiency and certainty in the selection of desired traits were needed, the use of biotechnology method such as marker assisted selection and anther culture may be appropriate.

The method to develop insect-resistant plant can be grouped into two, i.e. conventional and biotechnology. The conventional approaches use the whole living organism, while biotechnology approaches use cells or biomolecules to develop insect-resistant plants. The conventional method includes pure line selection, mass selection, hybridization (pedigree, bulk, single seed descent, backcross, recurrent selection). While biotechnology approach includes protoplast fusion, somaclonal variation, molecular assisted selection, and genetic transformation (Collin and Edwards, 1998; Mujeeb-Kazi and Sitch, 1989). For anther culture and embryo culture, at the beginning they used organism of F1 plants to produce pollen grain and wild and cultivated

species to produce embryo. However, since in the following steps they may be using cells culture, so we consider the anther culture and embryo culture as a biotechnology approach. A short brief about bio-technology approach is as follows.

Anther Culture

Anther or pollen grain could be cultured *in vitro* using artificial medium. On the artificial medium anther may form callus, shoot, root, and finally the entire plants. All the plant are haploid. This approach is possible to speed up the formation of homozygous population of insect-resistant plants.

Embryo Culture

Wild species are often more resistant to insects. Wide hybridization to transfer genes conferring insect-resistant from wild species to cultivated plants will produce abnormal interspecific hybrid embryos. The embryo can be rescued by culturing it on the nutrient medium to generate the entire plants. Several genes for insect resistance have been transferred from wild to cultivated germplasm (Table 1)

Protoplast Fusion

Protoplasts are plant cells that could be isolated by digested the wall enzymatically. The traits of resistant to the pest may be present in the one of two species that cannot be hybridized sexually. The two species may be formed hybrids through protoplasts fusion. The protoplasts may be cultured on artificial medium and some protoplasts will grow into entire plants.

The plants may be carried the resistant traits.

Somaclonal Variation

Somaclonal variation may be used to select insect-resistant variants. Insect-resistant somaclones can be selected through the following steps (1) calli or cell suspension derived from high yielding variety were grown for several or long term cycles, (2) the long-term cell lines were regenerated into plants, and (3) the regenerated plants were evaluated against target insects.

About 2000 plantlets of sugar cane were evaluated for resistance to the sugar cane borer under artificial infestation as well as natural infestation in field plots. Some somaclones showed resistance to sugar cane borer. The same method has been used to obtain somaclones of sorghum showed resistance to the fall armyworm.

Marker Assisted Selection

Nucleic acid probe, antibodies, or enzymes may be used as a marker assisted selection in breeding

program. Those probes may be used to determine the genetic constitutions of plants or plant extracts, includes the presence of resistant traits. The precise information provided by this method will increase the speed and certainty of selection progeny carry resistance traits in conventional plant breeding. The probes that tight linkage to the resistance traits have been identified (Table 2).

Genetic Transformation

Genetic transformation refers to the introduction of cloned DNA segments or genes from plants, bacteria, or animals into a new genetic background. The DNA segments or genes introduced into the protoplast or cell may confer resistance to insect. The genes may be delivered to the protoplast or cell by using one of the following method: Agrobacterium, biolistic, electro-poration, PEG, vortex, and micro-injection. Several resistant plants have been developed through genetic transformation (Table 3).

STRATEGIES TO DEVELOP INSECT-RESISTANT PLANTS

Table 1. Genes conferring insect resistance transferred from wild to cultivated species

Recipient	Alien donor	Trait
Bread wheat	<i>Secale cereale</i> <i>S. cereale</i>	Resistance to greenbug Resistance to Hessian fly
Rice	<i>Aegilops squarrosa</i> <i>Oryza officinalis</i> <i>O. officinalis</i> <i>O. australiensis</i> <i>O. minuta</i>	Resistance to Hessian fly Resistance to BPH Resistance to WBPH Resistance to BPH Resistance to BPH
Peanut	<i>Arachis monticola</i>	Resistance to chewing insect
Lettuce	<i>Lactuca virgata</i>	Resistance to Aphids
Cotton	<i>Gossypium armourianum</i>	Boll weevil, leafworm, bollworm

Source: Mujeeb-Kazi and Stich (1989)

Table 2. The probes for marker assisted selection

Crop	Trait of resistance	Genes	Probes
Rice	WBH	Wbph-1	RG146
	BPH	Bph-10(t)	RG457
	Gallmidge	Gm-2	RG476, RG776, RG 224
Mungbean	Bruchid beetle	-	PA 882
Wheat	Hessian fly	H-23	XksuH4, XksuG48(A)
		H-24	XcniBCD 457, Xcni CDO 482, Xksu G48 (B)

Source: Panda and Khush (1995)

Table 3. Resistant plants develop through genetic transformation

Plants	Traits
Alfalfa	Insect resistance
Cabbage	Insect resistance
Cotton	Resistance to bollworms and budworm (Bt toxin)
Eggplant	Insect resistance
Maize/Corn	Resistance to corn borer (Bt toxin)
Potato	Resistance to Colorado potato beetle (Bt toxin)
	Resistance to Potato tuber moths
Soybean	Insect resistance
Sugar cane	Insect resistance
Sweet potato	Insect resistance
Tobacco	Insect resistance
Tomato	Insect resistance
Rice	Insect resistance (Bt toxin)

Source: James (1999)

Sooner or later an insect-resistant plants will become susceptible due to the development of new biotypes. To anticipate the breakdown of an insect-resistant plants, various strategies should be adopted in developing those plants. One strategy is to prolong the useful life of insect-resistant plants. This can be realized by developing durable insect-resistant plant. The durable resistant plant is governed by polygenes. Durable resistance is also referred to poligenic resistance or horizontal resistance. The other strategy is to develop varieties with different genes conferring insect-resistant. This may be achieved by developing vertical resistant plants. The vertical resistant plant is governed by major genes. The more vertical resistant varieties with different genes is available, the more easy farmer access to new varieties to replace the previous varieties.

Durable Insect-resistant Plants

The level of resistance of durable insects-resistant plants is generally not very high. Because of this low selection pressure, the development of new biotypes is very slow. Polygenes resistance could be obtained from the landraces. Once crossing has been made between cultivated and landraces, we face to the difficulty in the selecting the desired segregants. This may be due to not all the polygenes or QTL from landraces are

transferred to cultivated and the level of resistance is diluted. The drawback is that the screening techniques currently available is only applicable to identify segregants with high level of resistance not segregants with low level of resistance.

Molecular-assisted selection offers great role to facilitate the development of durable insect-resistant plants. However, the first step should be done is to tagged the QTL for insect resistance with molecular markers. Molecular marker-based selection will assist in the accumulation of polygenes. QTLs for resistance to rice brown plant-hopper have been tagged with RFLP markers (Alam and Cohen, 1998).

Vertical Insect-resistant Plants

Vertical insect-resistant plants are more easier to develop since major genes are more easier to transfer from one variety to others. To anticipate the emergence of new insect biotypes that overcoming resistant plant, numerous vertical insect-resistant plants with different genes should be developed through conventional or biotechnology. Numerous rice varieties with different genes resistance to BPH have been produced by IRRI.

Insect-resistant transgenic plants developed through genetic transformation currently available is generally vertical resistance with the major genes, such Bt-gene. To

anticipate the development of new biotype overcoming Bt-plants, others resistant plants with different genes should be developed.

Pyramiding of major genes may be one of strategy to develop the longer useful life of resistance plants. This approach needs reliable method to determine the presence of the two genes. If bioassay method will be adopted, several insect biotypes should be available for this purpose. The use of molecular markers is very reliable method to diagnose the integration of the two genes in the plants. Pyramiding of major genes may be achieved through genetic transformation, for example pyramiding the two Bt genes or combining genes encoding a toxin and a gene encoding a repellent.

Tissue-specific Expression

The constitutive expression of resistance genes at all time and in the whole tissue may be caused a great selection pressure that leads to the development of new insect biotypes. This problem may be solved by expressing the resistance traits in a specific tissue, or at certain growth stages, or only in response to insect feeding. Promoters to regulate these expression are available.

CONCLUSION

Numerous insect-resistant plants have been developed through biotechnology. The use of biotechnology tools in breeding program will continue. Advances in biotechnology will accelerate the development of insect-resistant plants. The acceptability to biotechnology products may be greater a long with the increase in the understanding to biotechnology processes.

REFERENCES

- Alam, S.N. and M.B. Cohen. 1998.** Detection and analysis of QTLs for resistance to the brown plant-hopper, *Nilaparvata lugens*, in a double-haploid rice population. Theor. Appl. Genet. 97:1370-1379.
- Collin, H.A. and S. Edwards. 1998.** Plant cell culture. BIOS Scientific Publisher Limited. p. 158
- James, C. 1999.** Global review of commercialized transgenic crops. ISAAA Briefs 12:viii.
- James, C. and A.F. Krattiger. 1996.** Global review of the field testing and commercialization of transgenic plants. 1986 to 1995. The first decade of crop biotechnology. ISAAA Briefs 1:31.
- Mujeeb-Kazi, A. and L.A. Sitch. 1989.** Review of advances in plant biotechnology, 1985-88. 2nd International Symposium on Genetic Manipulation in Crops. Mexico, DF Mexico, and Manila, Philippines: CIMMYT and IRRI. p. 328.
- Panda, N. and G.S. Khush. 1995.** Host plant resistance to insects. IRRI. CAB International. p. 431
-